

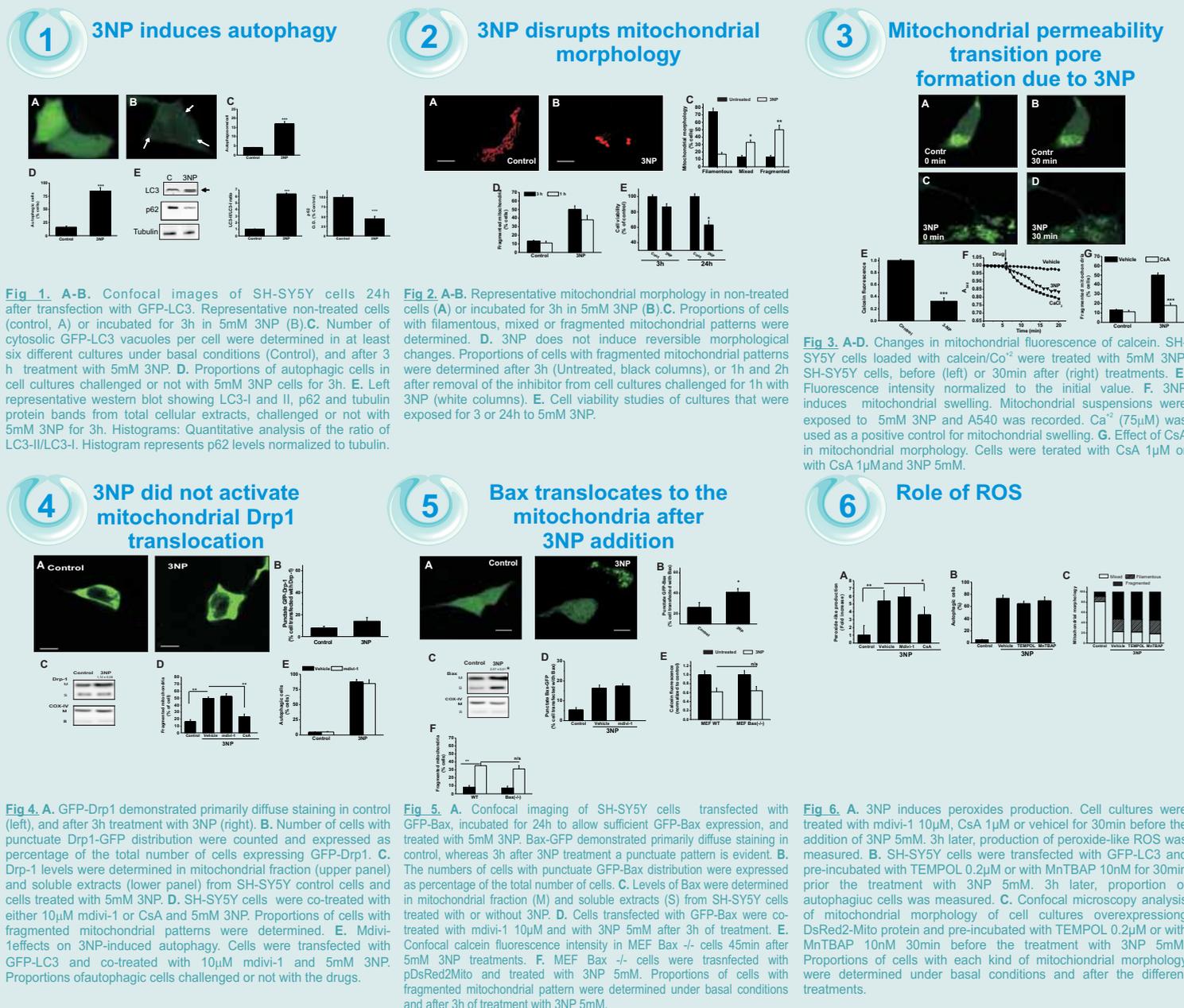
3-NITROPROPIONIC ACID INDUCES AUTOPHAGY BY MITOCHONDRIAL PERMEABILITY TRANSITION PORE RATHER THAN ACTIVATION OF THE MITOCHONDRIAL FISSION PATHWAY.

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Huntington's disease is a neurodegenerative process that has been associated with mitochondrial alterations including depletion in complex I activity. Drug inhibitors of complex I, such as 3-nitropropionic acid (3NP), are frequently used as pharmacological model to study the molecular and cellular pathways that are involved in this disease. Mitochondria are considered multifunctional organelles of changing morphology. Participation of the intrinsic apoptosis pathway, which is critically dependent on mitochondrial outer membrane permeabilization (MOMP), and the consequent release of cell-death mediating mitochondrial intermembrane space proteins, such as cytochrome c, has been described in this model. The knowledge of upstream modulators of mitochondrial dynamics is still not complete. Consistent with this, autophagy contributes to mitochondrial dysfunction-induced neurodegeneration and in vivo administration of 3NP activates autophagy. How 3NP regulates autophagy, and how this is related to mitochondrial morphology is an important question. Therefore, in the present study, we investigated the contribution of the mitochondrial-morphology pathway to autophagy activation as induced by 3-NP.



Statistics All data represent mean ± SEM of four independent experiments (*p < 0.05, **p < 0.001 and ***p < 0.001; t test versus control or one way ANOVA post hoc Tukey. Scale bars, 10 μm.

Conclusions SH-SY5Y cells presented a long, tubular and filamentous net of mitochondria. Upon 3NP treatment, mitochondria became dramatically shorter and rounder. Furthermore, 3NP induces the formation of mitochondrial permeability transition pore, both in cell cultures and in isolated liver mitochondria, and this process is sensitive to the presence of its inhibitor, cyclosporine A. Participation of the mitochondrial fission pathway is excluded because 3NP did not induce translocation of the DRP1 to the mitochondria. Also, inhibition of this GTPase, using mdivi-1, did not abrogate the observed mitochondrial alteration morphology. Finally, we ascertain the participation of reactive oxygen species in this neurodegenerative model, observing that scavenger drugs failed to prevent mitochondrial alterations while the presence of cyclosporine A, and not mdiv1, prevented ROS generation.

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